

## Invited Review

# Cellular Abnormalities and Emerging Biomarkers in Alcohol-Associated Liver Disease

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Alcohol-associated liver disease (AALD) is the third most common preventable cause for disease burden and mortality in the US. AALD, including alcoholic hepatitis (AH), contributes to half of admissions from decompensated liver disease and 20% of all liver transplants in the US. Peripheral blood cells contribute to systemic inflammation, oxidative stress, mitochondrial dysfunction, and fibrosis in AALD and AH. Alcohol dysregulates function of lymphocytes, neutrophils, monocytes, and tissue macrophages of the innate immune system. These alterations in turn can modulate adaptive immune responses. In this review, we describe these disruptive effects of alcohol on cells of the innate and adaptive immune system and focus on cellular-based emerging biomarkers on diagnosis and prognosis of patients with AALD and AH.

**Key words: Liver regeneration; Inflammation; Macrophage; Innate immunity; Hepatic injury; Gut–liver axis**

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## INTRODUCTION

Alcoholic liver disease is due to heavy alcohol consumption, which is typically defined as 3 drinks/day in men and 2 drinks/day in women for 5 years. As this amount of alcohol consumption may often exist with completely normal social and personal functioning of an individual, the word alcoholic may inappropriately stigmatize alcohol consumption behavior. Hence, there is increasing acceptance to change the terminology to alcohol-associated liver disease (AALD). Although, this change is still evolving, the authors in this review will use the term AALD when referring to alcoholic liver disease.

AALD is the third most common cause for chronic liver disease in the US after hepatitis C virus (HCV) infection and nonalcoholic fatty liver disease<sup>1</sup>. About two-thirds of the US population consumes alcohol at some point of their life, with 8%–10% engaging in heavy alcohol use<sup>2,3</sup>. The clinical spectrum of AALD varies from steatosis or fatty liver to more serious presentations such as steatohepatitis, fibrosis, cirrhosis, and hepatocellular carcinoma (HCC)<sup>4</sup>.

Advanced liver disease occurs in about 15%–20% of people with heavy alcohol use, with potential for cirrhosis and HCC<sup>4</sup>. Alcohol use remains the third most common preventable cause for any disease burden and mortality in the US after smoking and hypertension<sup>5</sup>. It also contributes to about 5% of global mortality and disease-adjusted life years lost, as AALD often occurs among relatively younger populations<sup>3</sup>.

A major contributor to this high mortality is from a unique clinical presentation of alcoholic hepatitis (AH), which presents with acute chronic liver failure among people with chronic and active use of alcohol in heavy amounts<sup>2,6</sup>. Disease awareness and better management have resulted in a decrease in AALD-related mortality in the general population over the last century from 20 to 25 to <10 per 100,000 between 1910 and 2006, respectively. However, this is far from 3 to 4 per 100,000, as the projected target for achievement by the WHO<sup>3</sup>. Further, outcome among patients with AH has not improved over the last five decades<sup>7</sup>. One of the reasons for not being

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able to achieve the targeted or expected decline in AALD mortality is the lack of effective and safe drugs for this potential lethal condition since the recognition of alcohol as a direct hepatotoxicant over six decades ago<sup>8</sup>. Moreover, advancement in the field has been hampered by the lack of animal models that accurately mimic the human phenotype of AALD and AH with potential for multi-organ failure, infections, and mortality<sup>9,10</sup>. However, recent interest in this area of research has led to the development of new models for study<sup>11</sup>.

Immune-activated peripheral blood cells contribute to the pathology of AALD and AH, including systemic inflammation, oxidative stress, mitochondrial function, and fibrosis. In the last few years, abnormalities in peripheral and tissue-resident blood cells in AALD have led to interesting and encouraging data on new biomarkers in the management of AALD. In this review, we focus on a) the effect of alcohol on innate and adaptive immunity; b) abnormalities in peripheral blood cells in stable cirrhosis and role of systemic inflammation in AALD and AH; c) mitochondrial dysfunction and cellular bioenergetics in AALD; and d) emerging biomarkers in the AALD field and their clinical utility throughout the course of AALD in diagnosis, predicting response to treatment, early diagnosis of infections, and determining prognosis.

### EFFECT OF ALCOHOL ON INNATE AND ADAPTIVE IMMUNITY

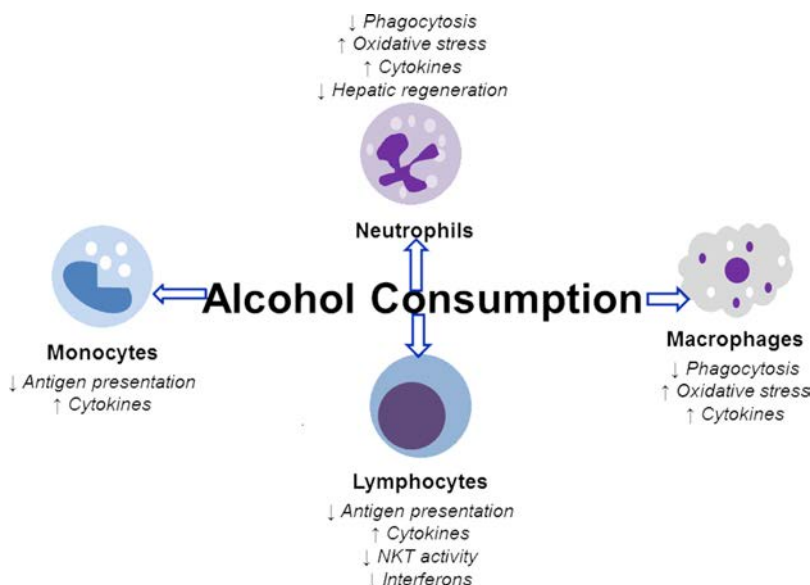
Alterations to the normal functioning of the gut–liver axis contribute to the pathology of AALD from multiple insults, including direct effects of alcohol on the innate

cells in the gastrointestinal tract<sup>12</sup>. Further, effects of alcohol on innate immunity can disrupt cross-talk between innate and adaptive immunity, resulting in inhibitory effects on adaptive immune responses. For example, alcohol consumption among patients with AALD induces bacterial overgrowth, dysbiosis of the gut microbiome with predominance of harmful bacteria, and increases gut permeability with translocation of gut bacteria across the portal circulation to hepatic parenchyma<sup>13</sup>. Toll-like receptors-4 (TLR-4) or pathogen-associated molecular pattern receptors on hepatic macrophages recognize these pathogens, resulting in inflammatory signaling and immune activation of peripheral blood cells<sup>14</sup>. This inflammatory signaling is mediated by NF- $\kappa$ B and, more importantly, IL-1 production from inflammasome activation<sup>13</sup>. The activation of macrophages via lipopolysaccharide (LPS) of the gut bacteria also mediates TLR-4 receptor activation<sup>15</sup>. The resulting liver injury releases danger-associated molecular patterns, which in turn stimulates TLR-4 perpetuating inflammatory signaling and pathology<sup>14</sup>.

Alcohol dysregulates key functions of most of the cells of the innate immune system including lymphocytes, neutrophils, monocytes, and tissue macrophages (Fig. 1).

#### Lymphocytes in AALD

Lymphocytes play an important role in the pathogenesis of AALD. In addition to being components of the innate immune system, these cells also provide adaptive immunity. As we understand, dendritic or antigen-presenting cells (APC) and natural killer (NK) cells are part of the immune system, while subsets of lymphocyte populations such as CD4, CD8, T regulatory cells,



**Figure 1.** Effects of alcohol consumption on peripheral blood cells.

B lymphocytes, and NK T cells are part of the adaptive immune system. Chronic alcohol consumption inhibits the activity and function of APC or dendritic cells, which is more pronounced in AALD compared to patients with cirrhosis from other etiologies<sup>16</sup>. Alcohol consumption activates T cells with production of cytokines and chemokines including upregulation of NF- $\kappa$ B-mediated inflammatory genes<sup>15</sup>. Alcohol inhibits the NK cells, which have anti-HCV and antifibrotic effects. While NK cells do not play much of a role in AALD per se, their inhibition by alcohol may explain a higher propensity and increased severity of disease in the presence of concomitant HCV among chronic alcoholics<sup>15</sup>.

NK T cells, a major population of hepatic lymphocytes, produce interferon (IFN) and IL-4 with other cytokines. NK T cells also protect against liver fibrosis by balancing liver cell damage and protective mediators such as IFN<sup>15</sup>. Dendritic cells promote hepatic injury by acting as APC to the T lymphocytes, with subsequent release of cytokines<sup>15</sup>. In another study, chronic alcohol ingestion activated CD4 and CD8 T lymphocytes in mice with AALD and without AALD compared to normal control mice<sup>17</sup>. These changes were associated with upregulation of CD80 and CD86, which are involved in T-cell-mediated APC interaction. These data suggest that chronic alcohol consumption causes immune activation of innate immunity and T cells<sup>17</sup>. Similar findings have been shown in humans with AH. For example, in one translational study, proportion of lymphocytes in the peripheral blood was reported to be increased in AH patients compared to healthy controls (35% vs. 25%), with this alteration persisting and remaining unchanged after 3 months of alcohol abstinence<sup>18</sup>. This increase occurred in T-cell populations with increase in CD4, CD8, and NK cells (CD3 and CD56), while the proportion of B cells (CD19/CD5) decreased<sup>18</sup>. Recent studies have also shown a role of upregulated programmed cell death 1 (PD1) and T-cell immunoglobulin and mucin domain-containing protein 3 (TIM-3) receptors, which inhibit host protective immunity. Antibodies to these receptors restored the function of cells with production of interferon, suggesting their potential utility in the treatment of AALD and severe AH<sup>19</sup>.

#### *Monocytes and Macrophages in AALD*

Like dendritic cells, monocytes also act as APC. Among patients with advanced liver disease and severe AH, monocytes fail to respond with production of TNF- $\alpha$ , which has been shown to be due to reduced human leucocyte antigen-D related (HLA-DR) and increased MER tyrosine kinase (MERTK) expression on cell surface<sup>20-22</sup>. A potential role of prostaglandin E2 has been postulated as inhibitory to the function of these cells<sup>23</sup>. These changes are associated with the inability of the monocytes

and tissue-resident macrophages to mount inflammatory responses against invading pathogens and antigens, resulting in immune paralysis with development of liver failure and secondary infections.

Kupffer cells comprise the majority of tissue-resident macrophages in the body and are activated by gut-derived endotoxin in response to alcohol. This activation in turn results in the release of cytokines and chemokines and sets the stage for inflammatory signaling, as well as reactive oxygen species production with oxidative stress<sup>24</sup>. The protective function of these cells is to clear the bacterial load and endotoxins and prevent infections. With the development of structural damage of the liver tissue in AALD and loss of tissue macrophages and Kupffer cells, patients remain at risk for bacterial infections, uncontrolled bacteremia, and sepsis<sup>25</sup>.

#### *Neutrophils in AALD*

Neutrophils play an important role against invading microorganisms, tissue trauma, or inflammatory signals. Recruitment of neutrophils occurs in response to acute liver injury or any systemic inflammatory signaling. Neutrophils accumulated in the hepatic microvasculature and sinusoids can transmigrate into the hepatic parenchyma if they receive a signal from distressed cells<sup>26</sup>. The main function of neutrophils recruited into the liver is to remove dead cells as a basis for regeneration, but neutrophils may also be involved in mediating hepatic inflammation and cell injury<sup>26</sup>. For example, recent studies demonstrate significant neutrophil accumulation in the liver following chronic plus binge ethanol feeding in mice with in vivo depletion of neutrophils by administration of anti-Ly6G antisera reducing liver injury<sup>27,28</sup>.

Neutrophil recruitment and activation are associated with increased proinflammatory cytokines such as IL-8 and TNF- $\alpha$  with decreased levels of anti-inflammatory cytokines such as IL-10<sup>29</sup>. After transmigration, neutrophils adhere to distressed hepatocytes through (2) integrins and ICAM-1 expressed on hepatocytes<sup>30</sup>. Neutrophil contact with hepatocytes mediates apoptosis<sup>31</sup>, as well as by oxidative killing via reactive oxygen species, leading to hepatocellular oncotic necrosis<sup>32-35</sup>. In one study of 35 AH patients, hepatocyte apoptotic index as measured by TUNEL staining was associated with severity of AH and Maddrey's discriminant function score, but not with the degree of neutrophilic infiltration on the liver tissue<sup>36</sup>.

Defects in phagocytosis and oxidative burst in neutrophils are transmissible by patient plasma to normal neutrophils, suggesting a role of soluble factors such as endotoxin and LPS<sup>37</sup>. There is a suggestion that administration of TLR-4 inhibitors may restore defects in oxidative burst, however not the phagocytic dysfunction of neutrophils, suggesting that TLR overexpression may be the result and not the cause of neutrophil activation<sup>37</sup>.

Recently, these defects in neutrophils have been shown to be due to a defect in myeloperoxidase release and the AKT/P38-MAPK pathway, which is salvageable with TLR-7/8 agonists, suggesting their potential therapeutic role<sup>38</sup>. Further, these defects have been linked to outcomes and with onset of infections and multiorgan failure<sup>39</sup>. Patients with cirrhosis do decompensate with infections as a precipitating factor, suggesting the presence of these neutrophil defects in stable cirrhosis patients<sup>40</sup>. In a study on 108 cirrhosis patients, neutrophil phagocytic dysfunction was associated with increased expression of TLR-4 receptors, resting respiratory burst, inflammatory cytokine levels, and phagocytic impairment. These defects were associated with the severity of cirrhosis and Child's class<sup>40</sup>. Although patients with compensated cirrhosis apparently had normal neutrophil function, a translational in vivo study showed phagocytic defect at the site of inflammation<sup>40</sup>.

The general purpose of early neutrophil infiltration is to remove dead or dying cells as a prerequisite for wound repair and regeneration. In fact, the degree of neutrophilic leucocytosis and the density of neutrophil infiltration on the hepatic tissue predict the outcome from an episode of AH among patients with alcohol-related cirrhosis and AALD<sup>41,42</sup>. In one study, levels of hepatic growth factor derived from recruited neutrophils into the liver were higher among 25 patients with AH compared to 20 nondrinker healthy controls and 20 patients with alcohol-related cirrhosis without AH<sup>43</sup>. Factors determining the protective role of neutrophils mediating hepatic regeneration versus its damaging effects on the liver tissue remain unknown. It is most likely that these defects are acquired during peripheral circulation of these cells through the bone marrow and reticuloendothelial system. Another speculation is that younger cells recruited from the bone marrow have better function and hepatic regenerative capacity. For example, use of growth factors such as granulocyte colony-stimulating factor in patients with acute-on-chronic liver failure and AH increases hepatic density of bone marrow-derived young CD34<sup>+</sup> cells, with benefits on hepatic regeneration, decreased occurrence of infections, improved liver disease, and better survival<sup>44</sup>.

### OXIDATIVE STRESS AND MITOCHONDRIAL DYSFUNCTION

Hepatic inflammation, oxidative stress, and mitochondrial dysfunction also contribute to the pathophysiology of AALD and AH<sup>26</sup>. Typically, mitochondrial dysfunction is defined as a partial loss of activity in the enzymatic components of oxidative phosphorylation, increases in generation of reactive oxygen species, and/or mutations and damage to mitochondrial DNA<sup>38,45,46</sup>. Mitochondrial dysfunction in AALD can be caused by numerous

factors following alcohol exposure, including inflammation from gut-derived bacterial endotoxin<sup>12</sup>, hepatic inflammation and apoptosis releasing damage-associated molecular patterns<sup>47</sup>, activation of Kupffer cells with nitric oxide synthase induction and nitrative stress<sup>48-51</sup>, and mitochondrial-mediated metabolism of alcohol via the CYP2E1 pathway of alcohol resulting in tissue hypoxia and enhanced oxidative stress<sup>50,52,53</sup>. The central role of mitochondrial dysfunction in AALD is also underscored by the recent finding that a mitochondrial-targeted antioxidant, MitoQ, prevents steatosis in an animal model of ethanol hepatotoxicity<sup>50</sup>. Similarly, treatment with S-adenosylmethionine and betaine prevents chronic alcohol-mediated impairments in mitochondrial function at multiple levels, including mtDNA damage, oxidant production, the proteome, and bioenergetics<sup>54</sup>. In addition, liver mitochondria after chronic alcohol administration are much more sensitive to nitrative stress from nitric oxide and reactive oxygen species-dependent protein inhibition<sup>49,55,56</sup>.

### *Bioenergetics in Peripheral Blood Cells as a Marker of Disease State and Mitochondrial Health*

Peripheral blood cells activated in a disease state can mediate processes involved in inflammation and cross-talk with body tissues. It is becoming clear that bioenergetics in peripheral blood cells may also reflect the disease state of peripheral organs and may be useful as markers of disease severity and outcomes in many diseases, including sepsis, diabetes mellitus, and neurodegenerative disorders<sup>57</sup>. In animal models, use of MitoQ and other mitochondrial-targeted antioxidants has been shown to be associated with reduction in steatosis, decreased nitrative stress, improved tissue hypoxia, and decrease in serum aminotransferase levels<sup>50,58</sup>. Thus, peripheral blood cells undergoing immune activation by the systemic inflammatory state in AALD and AH may be an appropriate setting to study mitochondrial health as biomarkers of disease state and prognosis.

The traditional approach to assess bioenergetics has been to isolate mitochondria from tissues or cells, and then assess biochemical function using measurement of oxygen consumption with added substrates or by performing individual enzymatic assays for each respiratory chain complex. The limitation of this strategy is that in vivo, extramitochondrial factors and compartments regulate the supply of substrates for oxidative phosphorylation, thus regulating mitochondrial metabolism<sup>59</sup>. With the advent of new high-throughput methods for the measurement of oxygen consumption in intact cells, bioenergetics can more readily be determined in clinical samples<sup>45,59</sup>. The resulting bioenergetics profile may then be useful for detecting subtle and early changes in bioenergetics function, which may then be predictive of

mitochondrial failure and development of disease<sup>25,57,59,60</sup>. It is also becoming evident that mitochondrial function in leukocytes is essential for normal immune responses<sup>27,50,56,61</sup>. This concept has precedent in other systemic and inflammatory disease states, where mitochondrial defects in monocytes and platelets have been described, such as diabetes mellitus, Alzheimer's disease, and septic shock<sup>44,61-64</sup>.

#### Assessment of Cellular Bioenergetics in Peripheral Blood Cells

To determine cellular bioenergetics, we and others use the XFe96 analyzer from Agilent Technologies (previously Seahorse Biosciences), which measures oxygen and protons (pH) in the extracellular environment of cell populations prepared using the techniques recently described<sup>45,57,65,66</sup>. For example, the system is capable of measuring 92 samples at a time (5 technical replicates and 18 different conditions; cell types or patients) and is equipped with four injection ports per well to allow for injection of a compound of interest or to add inhibitors that can aid in the elucidation of defects in individual cellular respiration pathways or enzymes<sup>57,65</sup>. Overall oxygen consumption rate (OCR) related to six key aspects of mitochondrial function can be measured as shown in Figure 2: a) basal mitochondrial respiration: this serves as baseline respiration; b) the proportion of basal activity that is linked to ATP production and level of proton leak (after injecting oligomycin in the medium): an increase in the proton leak is consistent with the mitochondria becoming less efficient; c) the maximal oxygen respiration (after injecting FCCP in the medium): a decrease in this parameter would be consistent with a deficit in

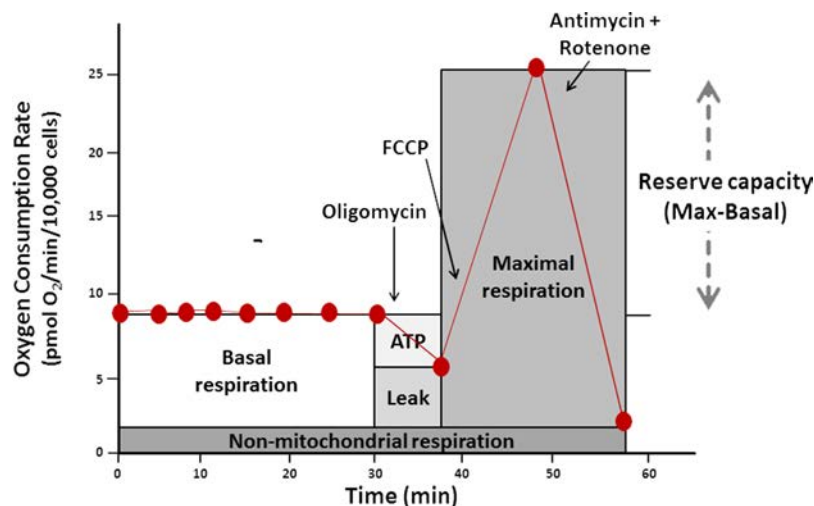
mitochondrial biogenesis, damaged mitochondrial DNA, and/or oxidative phosphorylation proteins; d) the reserve capacity (maximal minus basal respiration): determines the threshold at which bioenergetics dysfunction occurs (e.g., when reserve capacity is zero or negative, the cell cannot meet the bioenergetics demand through mitochondrial respiration and the bioenergetics threshold is exceeded<sup>66</sup>; and e) the level of nonmitochondrial oxygen consumption (after injecting antimycin in the medium): represents other oxygen-consuming processes in the cells, such as proinflammatory enzymes cyclooxygenases and lipoxygenases.

#### EMERGING BIOMARKERS IN AALD

Diagnosis of alcohol-related cirrhosis can be made accurately in most cases without liver biopsy and using clinical, biochemical, and imaging criteria<sup>67</sup>. However, some of the clinically unmet needs in the field of AALD are a) identifying subjects engaging in heavy alcohol consumption who are at risk for advanced liver disease, b) diagnosing AH without a liver biopsy, c) assessing response to specific treatment and personalized medicine, d) early diagnosis of infections, and e) prognosis and predicting outcomes of the disease state. These biomarkers can originate from blood or serum or plasma, urine, stool, breath, and cell-based assays. In the following sections, we highlight several potential new biomarkers in AALD with a summary provided in Table 1.

#### Biomarkers for Risk of Advanced Liver Disease

As advanced liver disease with cirrhosis and HCC develops in 15%–20% of people with heavy alcohol



**Figure 2.** Scheme of a typical extracellular flux (XF) experiment in which basal, ATP-linked, proton leak, nonmitochondrial, and maximal respiration, as well as reserve capacity, can be determined in cells following serial injections of oligomycin, FCCP, and antimycin + rotenone.

**Table 1.** Emerging Biomarkers in Alcohol-Associated Liver Disease (AALD) and Alcoholic Hepatitis (AH)

Biomarker	Comment
<b>Biomarkers for diagnosis of AH and severe spectrum of AALD</b>	
<i>PNPLA3</i> gene polymorphisms	<i>PNPLA3</i> polymorphisms associated with severe AH in AALD patients <sup>68,69</sup>
M1 macrophages	Transformation of hepatic and circulating macrophages to M1 phenotype in AH <sup>70,78,79</sup>
WBC and platelets	WBC and platelet counts >11 and >148,000, respectively, predict diagnosis of AH <sup>71</sup>
Trimethylamine (TMA)	Breath trimethylamine (TMA) levels to diagnose AH <sup>74</sup>
CCL-20	Higher CCL-20 in AH patients <sup>97</sup> (Fig. 3)
CK-18 fragments—M65 and M30	Higher CK-18 fragments and lower M30:M65 ratio in AH patients <sup>78,79,97</sup>
Monocyte bioenergetics	Decreased basal, ATP, and proton leak-linked oxygen consumption in AH patients <sup>57</sup>
<b>Biomarkers to assess treatment response and disease prognosis</b>	
Neutrophils	Neutrophil infiltration and neutrophilic leukocytosis predict better outcome in AH patients <sup>41</sup>
MELD and Lille score	Dynamic model with MELD at baseline and Lille score at 1 week of corticosteroid treatment is a better predictor of disease outcome from severe AH than either score alone <sup>82,90,92</sup>
SIRS and serum LPS	Multiorgan failure and mortality higher in AH patients with SIRS; LPS predictive of mortality <sup>93,94</sup>
Procalcitonin	Predictive of AH patients with SIRS and infection <sup>94</sup>
CK-18 fragments—M65 and M30	M30:M65 ratio predictive of 30-day outcome <sup>79</sup>
Gene signaling pathways	Upregulation of hepatic regenerative pathway genes associated with steroid response and good outcome; upregulation of inflammation pathway genes associated with nonresponse to steroids, progressive liver failure, and bad outcome <sup>86</sup>
Hepatic venous pressure gradient (HVPG)	Hepatic venous pressure gradient (HVPG) greater in nonsurvivors among severe AH patients (HVPG > 22 mmHg) <sup>98</sup>
Monocyte bioenergetics	Improved bioenergetics parameters in five of eight AH patients treated with corticosteroids <sup>57</sup>
<b>Biomarkers to predict development of infection in AALD and AH</b>	
Bacterial DNA	High circulating levels of bacterial DNA predictive of infection and infection-related mortality <sup>106</sup>
Oxidative burst	Decreased oxidative burst and reduced NADPH oxidase predictive of infection in AH <sup>106,109</sup>

consumption, other disease-modifying factors likely play a role in predisposing an individual to advanced liver disease<sup>4</sup>. There are emerging data on the *PNPLA3* GC and GG polymorphisms association with AH phenotype among patients with AALD<sup>68,69</sup>. For example, GC and GG polymorphisms of the *PNPLA3* gene compared to CC or wild-type gene predispose an individual to susceptibility of AALD and advanced and severe spectrum of the disease<sup>68</sup>. There is emerging data to show that the transformation of hepatic and peripheral circulating macrophages to proinflammatory phenotype or M1 macrophages with decreased immune activation and cytokine signaling characterizes the disease pathology of AH<sup>62,70</sup>. In this regard, studies designed to characterize the phenotype of peripheral macrophages are attractive as they might also serve as unique biomarkers to identify individuals likely to develop AH.

#### *Biomarkers for Noninvasive Diagnosis of AH*

As the clinical diagnosis of AH may be inaccurate in 20–30% of cases, either due to questionable history of alcohol consumption or confounding by the presence of

liver disease etiologies other than alcohol, liver biopsy remains the gold standard for its definitive diagnosis<sup>67</sup>. Liver biopsy being invasive is not without complications in these sick patients with liver and multiorgan failure with coagulopathy. Moreover, leucocytes and platelets respond to systemic inflammation with increases in their peripheral counts. For example, in a clinical study of suspected AH patients undergoing liver biopsy, a white cell count >11 and a platelet count of >148,000 had an accuracy of about 85%–90% for diagnosis of AH<sup>71</sup>. Interestingly, gut bacteria convert phosphatidylcholine, a component of dietary lipids to trimethylamine (or TMA), which further gets converted to TMA oxide by hepatic flavin monooxygenases<sup>72</sup>. Importantly, patients with chronic liver disease have reduced conversion of TMA to its oxide, which is augmented by intestinal bacterial overgrowth and dysbiosis induced by alcohol consumption<sup>73</sup>. In one study, breath levels of TMA were useful in stratifying AH diagnosis among patients presenting with decompensated AALD<sup>74</sup>. Alcohol exposure of the HepG2 cells also stimulates secretion of extracellular vesicles by hepatocytes, which then leads to

activation of macrophages, resulting in caspase 3 activation<sup>75</sup>. Cytokeratin-18 (CK-18), a main component of Mallory–Denk bodies, is released in response to hepatocyte injury in AH and is carried within the vesicles released from hepatocytes. Two components can be detected in the serum: one with total and the fragmented form (or M65) released in response to total cell death or necrosis, and the cleaved fraction (or M30), which is released in response to apoptosis<sup>76</sup>. Patients confirmed to have AH on transjugular liver biopsy have been shown to have higher levels of both the fractions of CK-18 and levels of chemokine CCL-20<sup>77,78</sup>. In another study, patients with AH compared to healthy controls and decompensated cirrhosis due to alcohol had lower M30:M65 ratio<sup>79</sup>. As systemic inflammation in AALD and AH suppresses mitochondrial function and bioenergetics, we examined the importance of studying this in peripheral monocytes in our prospective cohort consisting of 63 decompensated AALD patients, 28 with severe AH. Our preliminary data showed reduced basal, ATP linked, and proton leak-linked mitochondrial OCR in AH patients. In a logistic regression model, basal and ATP-mediated OCR predicted diagnosis of AH among decompensated AALD patients, after controlling for serum bilirubin and white blood cell count<sup>7</sup>. Further, in a subset of 32 (14 with severe AH) patients from this cohort, cytokine profile measured using the multiplex approach showed CCL-20 levels (mean±SEM) to be higher in AH patients compared to 18 patients with decompensated AALD without AH ( $1,256 \pm 57$  vs.  $434 \pm 36$  pg/ml,  $p=0.032$ ) (Fig. 3).

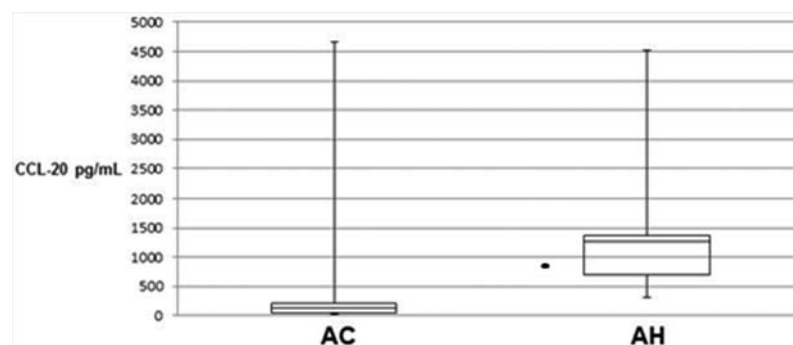
#### *Biomarkers for Predicting Response to Treatment*

Currently, corticosteroids are the only available therapeutic option for the treatment of severe AH<sup>80,81</sup>. However, these drugs are limited in a major way due to noneligibility in about 40% of AH patients, unpredictable nonresponse in about 40%, which can only be determined at 1 week after administration of these drugs, and risk

of complications and infections<sup>82</sup>. Further, the mortality benefit is limited to only about a month from the presentation, with no long-term survival benefit from these drugs<sup>83</sup>. Patients with hepatorenal syndrome also do not respond to corticosteroids, and it is recommended that the hepatorenal syndrome be treated first before initiation of corticosteroids<sup>80</sup>. Response to corticosteroids is inherently based on the immune activation in a patient with AH. This was shown in an in vitro study, where isolated lymphocytes from AH patients compared to those from healthy controls were less sensitive to inhibition of their proliferation by corticosteroids, and this was further reduced in nonresponders to corticosteroids compared to responders<sup>84</sup>. Thus, there remains a knowledge gap and critical need for simple biomarkers that will aid in predicting responsiveness to corticosteroids and also as a way to better personalize the medical therapy of severe AH patients.

Further, we have learned about the important beneficial aspects of hepatic and macrophage-derived TNF on the hepatic regenerative capacity from studies showing the detrimental effects of anti-TNF agents in patients with severe AH<sup>85</sup>. Further, neutrophil infiltration on the liver biopsy and neutrophilic leukocytosis in these patients are favorable markers for better prognosis and survival in severe AH<sup>41</sup>. In a recently reported study on severe AH patients from India and France, gene expression studies performed on the hepatic tissue showed that upregulation of genes involved in hepatic regenerative pathways is associated with responsiveness to steroids and better outcome<sup>86</sup>. Until these biomarkers are validated in multi-center studies with larger sample sizes, steroid responsiveness will continue to be determined using Lille score at 1 week or at least 4 days of steroid therapy<sup>87</sup>.

In our prospective cohort, of the eight AH patients treated with corticosteroids, repeat bioenergetics measurements on basal, ATP linked, and maximal mitochondrial OCR in the peripheral monocytes tended to improve in five responders (Lille score:  $<0.45$ ); however, this



**Figure 3.** Levels of chemokine CCL-20 in serum comparing 18 patients with alcoholic cirrhosis (AC) versus 14 patients with alcoholic hepatitis (AH).

tended to decrease or remain unchanged among three nonresponders<sup>7</sup>. Although corticosteroids predominantly have anti-inflammatory effects<sup>88</sup>, there is some evidence that these drugs may stimulate synthesis of mitochondrial proteins<sup>28,73</sup>. These data support the hypothesis that circulating leukocytes may serve as real-time sensors of mitochondrial function and liver disease pathology, as well as be sensitive to therapeutic intervention.

#### *Biomarkers for Assessing Prognosis of Disease*

Of the various scoring systems available to gauge disease severity and prognosis of patients with AH, traditionally modified discriminant function (mDF) index and model for end-stage disease (MELD) score are used in clinical practice<sup>89-91</sup>. Dynamic modeling using the MELD at baseline and Lille score at 1 week of corticosteroid treatment also appears to provide better prediction of outcomes than either of the two scores alone<sup>82,90,92</sup>. Of the clinical variables, hepatic encephalopathy and presence of systemic inflammatory response syndrome (SIRS) are the most consistent and reliable markers for disease severity and predicting outcome<sup>93,94</sup>. For example, SIRS is an accurate predictor for onset of renal failure in these patients, a major determinant of patient mortality<sup>95</sup>. Liver biopsy performed in about 15%–20% of these patients in the US mainly for diagnostic uncertainty also provides prognostic information<sup>41</sup>. It must be recognized that these scoring systems provide prognosis on a short-term basis, and the long-term outcomes in these patients are mainly dependent on their ability to remain sober after recovery from the episode of AH<sup>96</sup>, as about 30%–35% and 40%–50% of severe AH patients relapse to alcohol use within 6 months and 1 year, respectively<sup>83</sup>. Currently, there are no factors or variables that can accurately predict risk of relapse to alcohol use in this population.

Data are also emerging on the role of LPS and procalcitonin levels in predicting response to treatment and onset of multiorgan failure, a major cause of death in patients with severe AH<sup>94</sup>. Serum biomarkers of systemic inflammation especially extracellular vesicles, CCL-20, and CK-18 M65:M30 levels have shown promise in estimating disease outcome and prognosis in AH patients<sup>77-79</sup>. In one study, M30:M65 ratio was better than traditional scores of estimating disease severity of AH and its outcome at day 30 from presentation<sup>79</sup>. Progression of liver failure in AH and other liver diseases is a balance between progression of sterile inflammation and regenerative capacity of liver parenchyma. In this regard, levels of anti-inflammatory cytokines such as IL-10 or IL-22 and inflammatory cytokines and chemokines including CCL-20 have emerged as predictors of patient outcome<sup>97</sup>. Further, gene expression studies performed on liver tissue have shown upregulation of genes involved in hepatic regeneration among patients responsive to corticosteroids and good

outcome, and genes involved in inflammation signaling pathways upregulated in patients nonresponsive to treatment with progressive liver failure and bad outcome<sup>86</sup>. Invasive technique like hepatic venous pressure gradient measurement has also been shown to predict outcomes and disease course; however, this has not been used for this sole purpose in practice in this population<sup>98</sup>.

#### *Biomarkers for Early Diagnosis of Infections*

Infection is a common occurrence and follows patients with liver failure including those with severe AH<sup>99</sup>. Infections occur in about 30%–50% among hospitalized cirrhotic compared to 5%–7% of hospitalized patients from causes other than liver cirrhosis<sup>100,101</sup>. Abnormalities of innate and adaptive immunity especially in neutrophils among AALD patients make them susceptible to bacterial infections<sup>102</sup>. Also, immunocompromised state of cirrhosis and portal hypertension bypassing of gut bacteria and endotoxins to systemic circulation set the stage for development of infections in these patients<sup>103</sup>. Further, cytokine storm of ongoing inflammation and pathology in AH can also cause immune paralysis with susceptibility of these patients for multidrug-resistant and fungal nosocomial and hospital-acquired infections, which are less responsive to antibiotics, and may progress to development of septic shock and high mortality<sup>104,105</sup>. This risk increases further with treatment of these patients, especially with corticosteroids<sup>99,106</sup>. The accurate diagnosis of infections in these patients often becomes difficult, as features of SIRS are not accurate predictors of infection given the frequent presence of sterile inflammatory state in these patients<sup>107</sup>. Further, blood cultures can be negative in about 30%–40% of patients with bacterial sepsis, and there is often a lag time of many weeks before fungal culture results are available<sup>108</sup>. Over the last few years, biomarkers have emerged to predict bacterial infections in these patients, such as procalcitonin, levels of bacterial DNA, oxidative burst in monocyte and neutrophils, and extracellular vesicles in peripheral blood samples<sup>39,94,106,109</sup>.

#### *Limitations of the Existing Biomarkers*

Over the last decade or so, the field is widening and expanding on the identification of biomarkers at every stage of the disease of AH, including early diagnosis, estimating prognosis, identification of infections, and determining responsiveness to treatment. However, all these studies are from single centers with small sample size. Further, many of these tests and assays are expensive, involve a complex technique for measurement, and are unavailable at many centers and are not commercially available. Clearly, these encouraging data need further validation on larger samples from multicenter recruitment, before their routine application in routine clinical practice. The ongoing studies in the NIAAA-funded



consortia in the US are an ideal opportunity for joining hands with other centers and investigators to collect samples on well-characterized patients as a basis for the development of accurate, simple, validated, and reliable biomarkers for management of patients with AALD and severe AH.

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## REFERENCES

- Wong RJ, Aguilar M, Cheung R, Perumpail RB, Harrison SA, Younossi ZM, Ahmed A. Nonalcoholic steatohepatitis is the second leading etiology of liver disease among adults awaiting liver transplantation in the United States. *Gastroenterology* 2015;148(3):547–55.
- Singal AK, Kodali S, Vucovich LA, Darley-Usmar V, Schiano TD. Diagnosis and treatment of alcoholic hepatitis: A systematic review. *Alcohol Clin Exp Res*. 2016; 40(7):1390–402.
- Singal AK, Anand BS. Epidemiology of alcoholic liver disease. *Clin Liv Dis*. 2012;2:53–6.
- Gao B, Bataller R. Alcoholic liver disease: Pathogenesis and new therapeutic targets. *Gastroenterology* 2011; 141(5):1572–85.
- Bauer UE, Briss PA, Goodman RA, Bowman BA. Prevention of chronic disease in the 21st century: Elimination of the leading preventable causes of premature death and disability in the USA. *Lancet* 2014;384(9937):45–52.
- Lucey MR, Mathurin P, Morgan TR. Alcoholic hepatitis. *N Engl J Med*. 2009;360(26):2758–69.
- Hughes E, Hopkins LJ, Parker R. Survival from alcoholic hepatitis has not improved over time. *PLoS One* 2018; 13(2):e0192393.
- Lieber CS, DeCarli LM. An experimental model of alcohol feeding and liver injury in the baboon. *J Med Primatol*. 1974;3(3):153–63.
- Mandrekar P, Bataller R, Tsukamoto H, Gao B. Alcoholic hepatitis: Translational approaches to develop targeted therapies. *Hepatology* 2016;64(4):1343–55.
- Wilkin RJ, Lalor PF, Parker R, Newsome PN. Murine models of acute alcoholic hepatitis and their relevance to human disease. *Am J Pathol*. 2016;186(4):748–60.
- Shah VH. Alcoholic liver disease: The buzz may be gone, but the hangover remains. *Hepatology* 2010;51(5): 1483–4.
- Keshavarzian A, Holmes EW, Patel M, Iber F, Fields JZ, Pethkar S. Leaky gut in alcoholic cirrhosis: A possible mechanism for alcohol-induced liver damage. *Am J Gastroenterol*. 1999;94(1):200–7.
- Szabo G. Gut-liver axis in alcoholic liver disease. *Gastroenterology* 2015;148(1):30–6.
- Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell* 2006;124(4):783–801.
- Gao B, Seki E, Brenner DA, Friedman S, Cohen JI, Nagy L, Szabo G, Zakhari S. Innate immunity in alcoholic liver disease. *Am J Physiol Gastrointest Liver Physiol*. 2011;300(4):G516–25.
- Munoz L, Jose Borrero M, Ubeda M, Lario M, Diaz D, Frances R, Monserrat J, Pastor O, Aguado-Fraile E, Such J, and others. Interaction between intestinal dendritic cells and bacteria translocated from the gut in rats with cirrhosis. *Hepatology* 2012;56(5):1861–9.
- Cook RT, Zhu X, Coleman RA, Ballas ZK, Waldschmidt TJ, Ray NB, LaBrecque DR, Cook BL. T-cell activation after chronic ethanol ingestion in mice. *Alcohol* 2004; 33(3):175–81.
- Laso FJ, Madruga JI, Lopez A, Ciudad J, Alvarez-Mon M, San Miguel J, Orfao A. Abnormalities of peripheral blood T lymphocytes and natural killer cells in alcoholic hepatitis persist after a 3-month withdrawal period. *Alcohol Clin Exp Res*. 1997;21(4):672–6.
- Markwick LJ, Riva A, Ryan JM, Cooksley H, Palma E, Tranah TH, Manakkat Vijay GK, Vergis N, Thursz M, Evans A and others. Blockade of PD1 and TIM3 restores innate and adaptive immunity in patients with acute alcoholic hepatitis. *Gastroenterology* 2015;148(3): 590–602.
- Wasnuth HE, Kunz D, Yagmur E, Timmer-Stranghoner A, Vidacek D, Siewert E, Bach J, Geier A, Purucker EA, Gressner AM, and others. Patients with acute on chronic liver failure display “sepsis-like” immune paralysis. *J Hepatol*. 2005;42(2):195–201.
- Bernsmeier C, Pop OT, Singanayagam A, Triantafyllou E, Patel VC, Weston CJ, Curbishley S, Sadiq F, Vergis N, Khamri W, and others. Patients with acute-on-chronic liver failure have increased numbers of regulatory immune cells expressing the receptor tyrosine kinase MERTK. *Gastroenterology* 2015;148(3):603–615.
- Cabezón R, Carrera-Silva EA, Florez-Grau G, Errasti AE, Calderon-Gomez E, Lozano JJ, Espana C, Ricart E, Panes J, Rothlin CV, and others. MERTK as negative regulator of human T cell activation. *J Leukoc Biol*. 2015; 97(4):751–60.
- O’Brien AJ, Fullerton JN, Massey KA, Auld G, Sewell G, James S, Newson J, Karra E, Winstanley A, Alazawi W, and others. Immunosuppression in acutely decompensated cirrhosis is mediated by prostaglandin E2. *Nat Med*. 2014;20(5):518–23.
- Arteel G, Marsano L, Mendez C, Bentley F, McClain CJ. Advances in alcoholic liver disease. *Best Pract Res Clin Gastroenterol*. 2003;17(4):625–47.
- Helmy KY, Katschke KJ, Jr., Gorgani NN, Kljavin NM, Elliott JM, Diehl L, Scales SJ, Ghilardi N, van Lookeren Campagne M. CR1g: A macrophage complement receptor required for phagocytosis of circulating pathogens. *Cell* 2006;124(5):915–27.
- Ramaiah SK, Jaeschke H. Role of neutrophils in the pathogenesis of acute inflammatory liver injury. *Toxicol Pathol*. 2007;35(6):757–66.
- Bertola A, Park O, Gao B. Chronic plus binge ethanol feeding synergistically induces neutrophil infiltration and liver injury in mice: A critical role for E-selectin. *Hepatology* 2013;58(5):1814–23.
- Mathews S, Feng D, Maricic I, Ju C, Kumar V, Gao B. Invariant natural killer T cells contribute to chronic-plus-binge ethanol-mediated liver injury by promoting hepatic neutrophil infiltration. *Cell Mol Immunol*. 2016;13(2): 206–16.
- Taieb J, Mathurin P, Elbim C, Cluzel P, Arce-Vicioso M, Bernard B, Opolon P, Gougerot-Pocidalo MA, Poynard T, Chollet-Martin S. Blood neutrophil functions and cytokine release in severe alcoholic hepatitis: Effect of corticosteroids. *J Hepatol*. 2000;32(4):579–86.

30. Jaeschke H, Hasegawa T. Role of neutrophils in acute inflammatory liver injury. *Liver Int.* 2006;26(8):912–9.
31. Faouzi S, Burckhardt BE, Hanson JC, Campe CB, Schrum LW, Rippe RA, Maher JJ. Anti-Fas induces hepatic chemokines and promotes inflammation by an NF-kappa B-independent, caspase-3-dependent pathway. *J Biol Chem.* 2001;276(52):49077–82.
32. Gujral JS, Farhood A, Bajt ML, Jaeschke H. Neutrophils aggravate acute liver injury during obstructive cholestasis in bile duct-ligated mice. *Hepatology* 2003;38(2):355–63.
33. Ramaiah SK, Jaeschke H. Hepatic neutrophil infiltration in the pathogenesis of alcohol-induced liver injury. *Toxicol Mech Methods* 2007;17(7):431–40.
34. Jaeschke H. Involvement of Kupffer cells in the interaction between neutrophils and sinusoidal endothelial cells in rats. *Shock* 18(2):152–157, 2002. Comment in *Shock* 2003;19(4):394–5; author reply 395–6.
35. Jaeschke H, Ho YS, Fisher MA, Lawson JA, Farhood A. Glutathione peroxidase-deficient mice are more susceptible to neutrophil-mediated hepatic parenchymal cell injury during endotoxemia: Importance of an intracellular oxidant stress. *Hepatology* 1999;29(2):443–50.
36. Ziol M, Tepper M, Lohez M, Arcangeli G, Ganne N, Christidis C, Trinchet JC, Beaugrand M, Guillet JG, Guettier C. Clinical and biological relevance of hepatocyte apoptosis in alcoholic hepatitis. *J Hepatol.* 2001;34(2):254–60.
37. Stadlbauer V, Mookerjee RP, Wright GA, Davies NA, Jurgens G, Hallstrom S, Jalan R. Role of Toll-like receptors 2, 4, and 9 in mediating neutrophil dysfunction in alcoholic hepatitis. *Am J Physiol Gastrointest Liver Physiol.* 2009;296(1):G15–22.
38. Boussif A, Rolas L, Weiss E, Bouriche H, Moreau R, Perianin A. Impaired intracellular signaling, myeloperoxidase release and bactericidal activity of neutrophils from patients with alcoholic cirrhosis. *J Hepatol.* 2016;64(5):1041–8.
39. Mookerjee RP, Stadlbauer V, Lidder S, Wright GA, Hodges SJ, Davies NA, Jalan R. Neutrophil dysfunction in alcoholic hepatitis superimposed on cirrhosis is reversible and predicts the outcome. *Hepatology* 2007;46(3):831–40.
40. Tritto G, Bechlis Z, Stadlbauer V, Davies N, Frances R, Shah N, Mookerjee RP, Such J, Jalan R. Evidence of neutrophil functional defect despite inflammation in stable cirrhosis. *J Hepatol.* 2011;55(3):574–81.
41. Altamirano J, Miquel R, Katoonizadeh A, Abraldes JG, Duarte-Rojo A, Louvet A, Augustin S, Mookerjee RP, Michelena J, Smyrk TC and others. A histologic scoring system for prognosis of patients with alcoholic hepatitis. *Gastroenterology* 2014;146(5):1231–9.
42. Mathurin P, Duchatelle V, Ramond MJ, Degott C, Bedossa P, Erlinger S, Benhamou JP, Chaput JC, Rueff B, Poynard T. Survival and prognostic factors in patients with severe alcoholic hepatitis treated with prednisolone. *Gastroenterology* 1996;110(6):1847–53.
43. Taieb J, Delarche C, Paradis V, Mathurin P, Grenier A, Crestani B, Dehoux M, Thabut D, Gougerot-Pocidal MA, Poynard T, and others. Polymorphonuclear neutrophils are a source of hepatocyte growth factor in patients with severe alcoholic hepatitis. *J Hepatol.* 2002;36(3):342–8.
44. Kedarisetty CK, Anand L, Bhardwaj A, Bhadoria AS, Kumar G, Vyas AK, David P, Trehanpati N, Rastogi A, Bihari C, and others. Combination of granulocyte colony-stimulating factor and erythropoietin improves outcomes of patients with decompensated cirrhosis. *Gastroenterology* 2015;148(7):1362–70.
45. Hill BG, Benavides GA, Lancaster JR Jr, Ballinger S, Dell'Italia L, Jianhua Z, Darley-USmar VM. Integration of cellular bioenergetics with mitochondrial quality control and autophagy. *Biol Chem.* 2012;393(12):1485–512.
46. Brookes PS, Levonen AL, Shiva S, Sarti P, Darley-USmar VM. Mitochondria: Regulators of signal transduction by reactive oxygen and nitrogen species. *Free Radic Biol Med.* 2002;33(6):755–64.
47. Mantena SK, King AL, Andringa KK, Eccleston HB, Bailey SM. Mitochondrial dysfunction and oxidative stress in the pathogenesis of alcohol- and obesity-induced fatty liver diseases. *Free Radic Biol Med.* 2008;44(7):1259–72.
48. McKim SE, Gabele E, Isayama F, Lambert JC, Tucker LM, Wheeler MD, Connor HD, Mason RP, Doll MA, Hein DW, and others. Inducible nitric oxide synthase is required in alcohol-induced liver injury: Studies with knockout mice. *Gastroenterology* 2003;125(6):1834–44.
49. Venkatraman A, Shiva S, Wigley A, Ulasova E, Chhieng D, Bailey SM, Darley-USmar VM. The role of iNOS in alcohol-dependent hepatotoxicity and mitochondrial dysfunction in mice. *Hepatology* 2004;40(3):565–73.
50. Chacko BK, Srivastava A, Johnson MS, Benavides GA, Chang MJ, Ye Y, Jhala N, Murphy MP, Kalyanaraman B, Darley-USmar VM. Mitochondria-targeted ubiquinone (MitoQ) decreases ethanol-dependent micro and macro hepatosteatosis. *Hepatology* 2011;54(1):153–63.
51. Song BJ, Abdelmegeed MA, Henderson LE, Yoo SH, Wan J, Purohit V, Hardwick JP, Moon KH. Increased nitroxidative stress promotes mitochondrial dysfunction in alcoholic and nonalcoholic fatty liver disease. *Oxid Med Cell Longev.* 2013;2013:781050.
52. Hoek JB, Cahill A, Pastorino JG. Alcohol and mitochondria: A dysfunctional relationship. *Gastroenterology* 2002;122(7):2049–63.
53. Ingelman-Sundberg M, Ronis MJ, Lindros KO, Eliasson E, Zhukov A. Ethanol-inducible cytochrome P4502E1: Regulation, enzymology and molecular biology. *Alcohol Alcohol Suppl.* 1994;2:131–9.
54. Bailey SM, Robinson G, Pinner A, Chamlee L, Ulasova E, Pompilius M, Page GP, Chhieng D, Jhala N, Landar A, and others. S-adenosylmethionine prevents chronic alcohol-induced mitochondrial dysfunction in the rat liver. *Am J Physiol Gastrointest Liver Physiol.* 2006;291(5):G857–67.
55. Venkatraman A, Shiva S, Davis AJ, Bailey SM, Brookes PS, Darley-USmar VM. Chronic alcohol consumption increases the sensitivity of rat liver mitochondrial respiration to inhibition by nitric oxide. *Hepatology* 2003;38(1):141–7.
56. Zelickson BR, Benavides GA, Johnson MS, Chacko BK, Venkatraman A, Landar A, Betancourt AM, Bailey SM, Darley-USmar VM. Nitric oxide and hypoxia exacerbate alcohol-induced mitochondrial dysfunction in hepatocytes. *Biochim Biophys Acta* 2011;1807(12):1573–82.
57. Chacko BK, Kramer PA, Ravi S, Benavides GA, Mitchell T, Dranka BP, Ferrick D, Singal AK, Ballinger SW, Bailey SM, and others. The Bioenergetic Health Index: A new concept in mitochondrial translational research. *Clin Sci. (Lond)* 2014;127(6):367–73.

58. Zhang P, Qiang X, Zhang M, Ma D, Zhao Z, Zhou C, Liu X, Li R, Chen H, Zhang Y. Demethyleneberberine, a natural mitochondria-targeted antioxidant, inhibits mitochondrial dysfunction, oxidative stress, and steatosis in alcoholic liver disease mouse model. *J Pharmacol Exp Ther.* 2015;352(1):139–47.
59. Brand MD, Nicholls DG. Assessing mitochondrial dysfunction in cells. *Biochem J.* 2011;435(2):297–312.
60. Dranka BP, Benavides GA, Diers AR, Giordano S, Zelickson BR, Reily C, Zou L, Chatham JC, Hill BG, Zhang J, and others. Assessing bioenergetic function in response to oxidative stress by metabolic profiling. *Free Radic Biol Med.* 2011;51(9):1621–35.
61. Widlansky ME, Wang J, Shenouda SM, Hagen TM, Smith AR, Kizhakekuttu TJ, Kluge MA, Weihrauch D, Gutterman DD, Vita JA. Altered mitochondrial membrane potential, mass, and morphology in the mononuclear cells of humans with type 2 diabetes. *Transl Res.* 2010;156(1):15–25.
62. Woolbright BL, Jaeschke H. Alcoholic hepatitis: Lost in translation. *J Clin Transl Hepatol.* 2018;6(1):89–96.
63. Japiassu AM, Santiago AP, d'Avila JC, Garcia-Souza LF, Galina A, Castro Faria-Neto HC, Bozza FA, Oliveira MF. Bioenergetic failure of human peripheral blood monocytes in patients with septic shock is mediated by reduced F1Fo adenosine-5'-triphosphate synthase activity. *Crit Care Med.* 2011;39(5):1056–63.
64. Ienco EC, Simoncini C, Orsucci D, Petrucci L, Filosto M, Mancuso M, Siciliano G. May “mitochondrial eve” and mitochondrial haplogroups play a role in neurodegeneration and Alzheimer’s disease? *Int J Alzheimers Dis.* 2011;2011:709061.
65. Chacko BK, Kramer PA, Ravi S, Johnson MS, Hardy RW, Ballinger SW, Darley-Usmar VM. Methods for defining distinct bioenergetic profiles in platelets, lymphocytes, monocytes, and neutrophils, and the oxidative burst from human blood. *Lab Invest.* 2013;93(6):690–700.
66. Dranka BP, Hill BG, Darley-Usmar VM. Mitochondrial reserve capacity in endothelial cells: The impact of nitric oxide and reactive oxygen species. *Free Radic Biol Med.* 2010;48(7):905–14.
67. Crabb DW, Bataller R, Chalasani NP, Kamath PS, Lucey M, Mathurin P, McClain C, McCullough A, Mitchell MC, Morgan TR, and others. Standard definitions and common data elements for clinical trials in patients with alcoholic hepatitis: Recommendation from the NIAAA Alcoholic Hepatitis Consortia. *Gastroenterology* 2016;150(4):785–90.
68. Salameh H, Raff E, Erwin A, Seth D, Nischalke HD, Falleti E, Burza MA, Leathert J, Romeo S, Molinaro A, and others. PNPLA3 gene polymorphism is associated with predisposition to and severity of alcoholic liver disease. *Am J Gastroenterol.* 2015;110(6):846–56.
69. Liangpunsakul S, Beaudoin JJ, Shah VH, Puri P, Sanyal AJ, Kamath PS, Lourens SG, Tang Q, Katz BP, Crabb DW, and others. Interaction between the patatin-like phospholipase domain-containing protein 3 genotype and coffee drinking and the risk for acute alcoholic hepatitis. *Hepatol Commun.* 2018;2(1):29–34.
70. Wan J, Benkdane M, Teixeira-Clerc F, Bonnafous S, Louvet A, Lafdil F, Pecker F, Tran A, Gual P, Mallat A, and others. M2 Kupffer cells promote M1 Kupffer cell apoptosis: A protective mechanism against alcoholic and nonalcoholic fatty liver disease. *Hepatology* 2014;59(1):130–42.
71. Hardy T, Wells C, Kendrick S, Hudson M, Day CP, Burt AD, Masson S, Stewart SF. White cell count and platelet count associate with histological alcoholic hepatitis in jaundiced harmful drinkers. *BMC Gastroenterol.* 2013;13(1):55.
72. Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, Feldstein AE, Britt EB, Fu X, Chung YM, and others. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* 2011;472(7341):57–63.
73. Velasquez MT, Ramezani A, Manal A, Raj DS. Trimethylamine N-oxide: The good, the bad and the unknown. *Toxins (Basel)* 2016;8(11).
74. Hanouneh IA, Zein NN, Cikach F, Dababneh L, Grove D, Alkhouri N, Lopez R, Dweik RA. The breathprints in patients with liver disease identify novel breath biomarkers in alcoholic hepatitis. *Clin Gastroenterol Hepatol.* 2014;12:516–23.
75. Verma VK, Li H, Wang R, Hirsova P, Mushref M, Liu Y, Cao S, Contreras PC, Malhi H, Kamath PS, and others. Alcohol stimulates macrophage activation through caspase-dependent hepatocyte derived release of CD40L containing extracellular vesicles. *J Hepatol.* 2016;64(3):651–60.
76. Cave M, Falkner KC, Ray M, Joshi-Barve S, Brock G, Khan R, Bon Homme M, McClain CJ. Toxicant-associated steatohepatitis in vinyl chloride workers. *Hepatology* 2010;51(2):474–81.
77. Affo S, Morales-Ibanez O, Rodrigo-Torres D, Altamirano J, Blaya D, Dapito DH, Millan C, Coll M, Caviglia JM, Arroyo V, and others. CCL20 mediates lipopolysaccharide induced liver injury and is a potential driver of inflammation and fibrosis in alcoholic hepatitis. *Gut* 2014;63(11):1782–92.
78. Bissonnette J, Altamirano J, Devue C, Roux O, Payance A, Lebec D, Bedossa P, Valla D, Durand F, Ait Oufella H, and others. A prospective study of the utility of plasma biomarkers to diagnose alcoholic hepatitis. *Hepatology* 2017;66:555–63.
79. Woolbright BL, Bridges BW, Dunn W, Olson JC, Weinman SA, Jaeschke H. Cell death and prognosis of mortality in alcoholic hepatitis patients using plasma keratin-18. *Gene Expr.* 2017;17(4):301–12.
80. Singal AK, Bataller R, Ahn J, Kamath PS, Shah VH. ACG clinical guideline: Alcoholic liver disease. *Am J Gastroenterol.* 2018;113(2):175–94.
81. European Association for the Study of L. EASL clinical practical guidelines: Management of alcoholic liver disease. *J Hepatol.* 2012;57(2):399–420.
82. Louvet A, Naveau S, Abdelnour M, Ramond MJ, Diaz E, Fartoux L, Dharancy S, Texier F, Hollebecque A, Serfaty L, and others. The Lille model: A new tool for therapeutic strategy in patients with severe alcoholic hepatitis treated with steroids. *Hepatology* 2007;45(6):1348–54.
83. Thursz MR, Forrest EH, Ryder S, investigators S. Prednisolone or pentoxifylline for alcoholic hepatitis. *N Engl J Med.* 2015;373(3):282–3.
84. Kendrick SF, Henderson E, Palmer J, Jones DE, Day CP. Theophylline improves steroid sensitivity in acute alcoholic hepatitis. *Hepatology* 2010;52(1):126–31.
85. Boetticher NC, Peine CJ, Kwo P, Abrams GA, Patel T, Aqel B, Boardman L, Gores GJ, Harmsen WS, McClain CJ, and others. A randomized, double-blinded, placebo-controlled multicenter trial of etanercept in the treatment

- of alcoholic hepatitis. *Gastroenterology* 2008;135(6):1953–60.
86. Sharma S, Maras JS, Das S, Hussain S, Mishra AK, Shasthry SM, Sharma CB, Weiss E, Elkrief L, Rautou PE, and others. Pre-therapy liver transcriptome landscape in Indian and French patients with severe alcoholic hepatitis and steroid responsiveness. *Sci Rep.* 2017;7(1):6816.
  87. Garcia-Saenz-de-Sicilia M, Duvoor C, Altamirano J, Chavez-Araujo R, Prado V, de Lourdes Candolo-Martinelli A, Holanda-Almeida P, Becerra-Martins-de-Oliveira B, Fernandez-de-Almeida S, Bataller R, and others. A day-4 Lille model predicts response to corticosteroids and mortality in severe alcoholic hepatitis. *Am J Gastroenterol.* 2017;112(2):306–15.
  88. Hamdi H, Bigorgne A, Naveau S, Balian A, Bouchet-Delbos L, Cassard-Doulcier AM, Maillot MC, Durand-Gasselin I, Prevot S, Delaveaucoupet J, and others. Glucocorticoid-induced leucine zipper: A key protein in the sensitization of monocytes to lipopolysaccharide in alcoholic hepatitis. *Hepatology* 2007;46(6):1986–92.
  89. Singal AK, Shah VH. Alcoholic hepatitis: Prognostic models and treatment. *Gastroenterol Clin North Am.* 2011;40(3):611–39.
  90. Dunn W, Jamil LH, Brown LS, Wiesner RH, Kim WR, Menon KV, Malinchoc M, Kamath PS, Shah V. MELD accurately predicts mortality in patients with alcoholic hepatitis. *Hepatology* 2005;41(2):353–8.
  91. Maddrey WC, Boitnott JK, Bedine MS, Weber FL Jr, Mezey E, White RI Jr. Corticosteroid therapy of alcoholic hepatitis. *Gastroenterology* 1978;75(2):193–9.
  92. Louvet A, Labreuche J, Artru F, Boursier J, Kim DJ, O'Grady J, Trepo E, Nahon P, Ganne-Carrie N, Naveau S, and others. Combining data from liver disease scoring systems better predicts outcomes of patients with alcoholic hepatitis. *Gastroenterology* 2015;149(2):398–406.
  93. Ravi S, Bade KS, Hasanin M, Singal AK. Ammonia level at admission predicts in-hospital mortality for patients with alcoholic hepatitis. *Gastroenterol Rep. (Oxf)* 2017; 5:232–6.
  94. Michelena J, Altamirano J, Abalde JG, Affo S, Morales-Ibanez O, Sancho-Bru P, Dominguez M, Garcia-Pagan JC, Fernandez J, Arroyo V, and others. Systemic inflammatory response and serum lipopolysaccharide levels predict multiple organ failure and death in alcoholic hepatitis. *Hepatology* 2015;62(3):762–72.
  95. Altamirano J, Fagundes C, Dominguez M, Garcia E, Michelena J, Cardenas A, Guevara M, Pereira G, Torres-Vigil K, Arroyo V, and others. Acute kidney injury is an early predictor of mortality for patients with alcoholic hepatitis. *Clin Gastroenterol Hepatol.* 2012;10(1):65–71.
  96. Louvet A, Labreuche J, Artru F, Bouthors A, Rolland B, Saffers P, Lollivier J, Lemaitre E, Dharancy S, Lassailly G, and others. Main drivers of outcome differ between short and long-term in severe alcoholic hepatitis: A prospective study. *Hepatology* 2017;66:1464–73.
  97. Affo S, Morales-Ibanez O, Rodrigo-Torres D, Altamirano J, Blaya D, Dapito DH, Millan C, Coll M, Caviglia JM, Arroyo V, and others. CCL20 mediates lipopolysaccharide induced liver injury and is a potential driver of inflammation and fibrosis in alcoholic hepatitis. *Gut* 2014;63:1782–92.
  98. Rincon D, Lo Iacono O, Ripoll C, Gomez-Camarero J, Salcedo M, Catalina MV, Hernando A, Clemente G, Matilla A, Nunez O, and others. Prognostic value of hepatic venous pressure gradient for in-hospital mortality of patients with severe acute alcoholic hepatitis. *Aliment Pharmacol Ther.* 2007;25(7):841–8.
  99. Singal AK, Shah VH, Kamath PS. Infection in severe alcoholic hepatitis: Yet another piece in the puzzle. *Gastroenterology* 2017;152(5):938–40.
  100. Caly WR, Strauss E. A prospective study of bacterial infections in patients with cirrhosis. *J Hepatol.* 1993;18(3):353–8.
  101. Navasa M, Fernandez J, Rodes J. Bacterial infections in liver cirrhosis. *Ital J Gastroenterol Hepatol.* 1999;31(7):616–25.
  102. Cortese MM, Wolff M, Almeida-Hill J, Reid R, Ketcham J, Santosham M. High incidence rates of invasive pneumococcal disease in the White Mountain Apache population. *Arch Intern Med.* 1992;152(11):2277–82.
  103. Bonnel AR, Bunchorntavakul C, Reddy KR. Immune dysfunction and infections in patients with cirrhosis. *Clin Gastroenterol Hepatol.* 2011;9(9):727–38.
  104. Hmoud BS, Patel K, Bataller R, Singal AK. Corticosteroids and occurrence of and mortality from infections in severe alcoholic hepatitis: A meta-analysis of randomized trials. *Liver Int.* 2016;36(5):721–8.
  105. Bajaj JS, O'Leary JG, Reddy KR, Wong F, Olson JC, Subramanian RM, Brown G, Noble NA, Thacker LR, Kamath PS, and others. Second infections independently increase mortality in hospitalized patients with cirrhosis: The North American consortium for the study of end-stage liver disease (NACSELD) experience. *Hepatology* 2012;56(6):2328–35.
  106. Vergis N, Atkinson SR, Knapp S, Maurice J, Allison M, Austin A, Forrest EH, Masson S, McCune A, Patch D, and others. In patients with severe alcoholic hepatitis, prednisolone increases susceptibility to infection and infection-related mortality, and is associated with high circulating levels of bacterial DNA. *Gastroenterology* 2017;152(5):1068–77.
  107. Mookerjee RP, Lackner C, Stauber R, Stadlbauer V, Deheragoda M, Aigelsreiter A, Jalan R. The role of liver biopsy in the diagnosis and prognosis of patients with acute deterioration of alcoholic cirrhosis. *J Hepatol.* 2011; 55(5):1103–11.
  108. Singal AK, Salameh H, Kamath PS. Prevalence and in-hospital mortality trends of infections among patients with cirrhosis: A nationwide study of hospitalised patients in the United States. *Aliment Pharmacol Ther.* 2014;40(1):105–12.
  109. Vergis N, Khamri W, Beale K, Sadiq F, Aletrari MO, Moore C, Atkinson SR, Bernsmeier C, Possamai LA, Petts G and others. Defective monocyte oxidative burst predicts infection in alcoholic hepatitis and is associated with reduced expression of NADPH oxidase. *Gut* 2017; 66(3):519–29.