

## ***Tcea3* Regulates the Vascular Differentiation Potential of Mouse Embryonic Stem Cells**

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*Tcea3* is present in high concentrations in mouse embryonic stem cells (mESCs) and functions to activate *Lefty1*, a negative regulator of Nodal signaling. The Nodal pathway has numerous biological activities, including mesoderm induction and patterning in early embryogenesis. Here, we demonstrate that the suppression of *Tcea3* in mESCs shifts the cells from pluripotency into enhanced mesoderm development. Vascular endothelial growth factor A (VEGFA) and VEGFC, major transcription factors that regulate vasculogenesis, are activated in *Tcea3* knocked down (*Tcea3* KD) mESCs. Moreover, differentiating *Tcea3* KD mESCs have perturbed gene expression profiles with suppressed ectoderm and activated mesoderm lineage markers. Most early differentiating *Tcea3* KD cells expressed Brachyury-T, a mesoderm marker, whereas control cells did not express the gene. Finally, development of chimeric embryos that included *Tcea3* KD mESCs was perturbed.

Key words: *Tcea3*; Vasculogenesis; Mouse embryonic stem cells (mESCs)

### INTRODUCTION

Mouse embryonic stem cells (mESCs) derived from the inner cell mass (ICM) of peri-implantation blastocysts can proliferate indefinitely and commit to differentiation into all cell types in vitro and in vivo (1). Therefore, the study of ESCs—especially mESCs—has provided valuable insights into early embryogenesis in mammals (2). Extrinsic stimuli, such as exogenously imposed or autonomously produced cytokines, play critical roles in ESC fate determination by triggering distinct signaling cascades (3). Among these signals, Activin/Nodal signaling is recognized as a pivotal regulatory cue in instructing the fate of ESCs (4). Smad2-dependent Activin/Nodal signaling is dispensable for the self-renewal maintenance of mESCs, but is required for differentiation into various types of cells, especially mesoderm (5–7). Phospho-Smad2 activated by Activin/Nodal directly regulates the expression of a cohort of developmental regulators, such as *Tapbp*, to induce meso-endoderm differentiation (5).

The development of the vascular system is one of the earliest events in embryogenesis because the growing embryo needs a transportation system to supply oxygen and nutrients. Vascular development begins before the initiation

of embryonic circulation by producing endothelial cell progenitors, which are assembled together to form an early primitive vessel network (8). Genetic deletion studies in mice revealed that activation of the vascular endothelial growth factor (VEGF) pathway is essential for the generation of the ESC lineage and the vasculature (9,10).

*Tcea3* is an isoform of the transcription elongation factor TFIIS (11). Previously, we reported that *Tcea3* is highly enriched in mouse ESCs and inhibits Smad2 phosphorylation by activating the expression of *Lefty1*, a negative regulator of Nodal signaling (12). Like Activin/Nodal signaling, *Tcea3* is also dispensable for self-renewal maintenance, but overexpression of *Tcea3* impairs the in vitro differentiation capacity. Suppression of *Tcea3* leads to enhanced expression of meso-endoderm markers during in vitro differentiation. In addition, mesoderm marker genes are downregulated in *Tcea3*-overexpressing mESCs under self-renewing culture conditions, suggesting that *Tcea3* functions to regulate lineage differentiation of mESCs by modulating Smad2 phosphorylation (12). However, the role of *Tcea3* in vasculogenesis of ESCs has not been examined. We report here that *Tcea3* suppression enhances the vascular development potential of mESCs. We examined whether transcription

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factors regulating vascular development are activated in *Tcea3* knocked-down (*Tcea3* KD) mESCs. We also examined whether *Tcea3* KD mESCs differentiate more rapidly under endothelial differentiation culture conditions. Finally, we confirmed that chimeric embryos developed from *Tcea3* KD mESCs had highly vascularized skin at embryonic day 13.5 (E13.5). These findings support the importance of *Tcea3* expression during vasculogenesis by ESCs.

## MATERIALS AND METHODS

### Cell Culture

J1 mESCs (catalog No. SCRC-1010) were purchased from ATCC (www.atcc.org) and maintained as described previously (13). Briefly, mESCs were maintained on 0.1% gelatin-coated dishes in ESC medium, which consists of Dulbecco's modified Eagle's medium (DMEM, Gibco Invitrogen, Carlsbad, CA, USA) supplemented with 10% horse serum (Gibco Invitrogen), 2 mM glutamine, penicillin (100 U/ml), streptomycin (100 µg/ml) (Gibco Invitrogen), 1 × nonessential amino acids (Life Technologies), 0.1 mM 2-mercaptoethanol (Sigma-Aldrich, St. Louis, MO, USA), and LIF (1,000 U/ml) (Chemicon, Temecula, CA, USA). Human umbilical vein endothelial cells (HUVECs) were obtained from Modern Cell & Tissue Technologies, Inc. (Seoul, Korea) and cultured in gelatin-coated plates with EGM-2 medium and supplements (Lonza, CC-3162) in the presence of 5% CO<sub>2</sub> at 37°C.

### Differentiation of ESCs

To induce spontaneous differentiation, mESCs were cultured in LIF-deficient ESC medium. The medium was changed every 2 days for mESC culture or differentiation. To confirm differentiation, alkaline phosphatase activity was measured by using the EnzoLyte™ pNPP Alkaline Phosphatase Assay kit (AnaSpec, catalog No. 71230), according to the manufacturer's instructions.

For endothelial differentiation, mESCs and stable knockdown cells of *Tcea3* were seeded at 2 × 10<sup>5</sup> cells in a 0.1% gelatin-coated 60-mm plate. The following day, the cells were used for endothelial differentiation, which consisted of two steps. First, the ESC culture medium was changed to a LIF-deficient ESC medium containing VEGF (50 ng/ml) and basic fibroblast growth factor (bFGF) (20 ng/ml) and the cells were incubated for 48 h. Then, the differentiation medium was changed to EGM-2 medium with supplements (Lonza, CC-3162) for the next 5 days. After endothelial differentiation of mESCs, tube formation assays were performed to evaluate vessel formation by the differentiated endothelial cells. The differentiated cells and HUVECs were seeded at 3 × 10<sup>4</sup> cells per well in 96-well plates on Matrigel (BD Biosciences, MA) that was previously polymerized for 30 min at 37°C. Changes in cell morphology were observed and photographed 24 h after seeding.

### Genetic Modification of mESCs

A shRNA plasmid targeting mouse *Tcea3* was purchased (RMM3981-97073145, Open Biosystems, Huntsville, AL, USA) to generate stable knockdown cell lines of *Tcea3*. shRNA plasmids were transfected into mESCs with Lipofectamine 2000 (Invitrogen) and stably transfected lines were established according to the manufacturer's instructions.

### Microarrays

Total RNA was extracted using TRIzol (Invitrogen), and biotinylated cRNA was prepared from 0.55 µg of total RNA using the Illumina TotalPrep RNA Amplification Kit (Ambion, Austin, TX, USA) according to the manufacturer's instructions. Following fragmentation, 1.5 µg of cRNA was hybridized to the Illumina Mouse WG-6 Expression Beadchip according to the manufacturer's instructions (Illumina, Inc., San Diego, CA, USA). Arrays were scanned with an Illumina Bead Array Reader Confocal Scanner according to the manufacturer's instructions. Array data processing and analysis were performed using Illumina BeadStudio v3.1.3 (Gene Expression Module v3.3.8).

### Generation and Analysis of Chimeric Embryos

To determine the in vivo developmental potential of *Tcea3* KD mESCs, chimeric embryos were made with GFP-expressing mESCs using standard ES cell transfer procedures for chimera production (Macrogen, Seoul, Korea). Typically, 8 to 12 GFP-expressing mESCs were injected into the cavity of a blastocyst from C57BL/6 mice. A total of 18–19 injected blastocysts were transferred into the uterus of a pseudopregnant ICR female at E2.5. Embryos were dissected at E13.5, and the contribution of injected mESCs was determined by the intensity of the GFP signal. For further histological analysis, embryos were embedded in paraffin using standard methods (14). Sections were cut at 7-µm thickness and stained with hematoxylin and eosin (H&E).

### Immunocytochemistry

J1 mESCs and *Tcea3* KD cells were cultured on gelatin-coated cover slips. The cells were washed twice with PBS and fixed with 4% paraformaldehyde for 10 min. Next, the cells were incubated in blocking buffer (5% bovine serum albumin in PBS) at room temperature for 1 h and then in anti-Brachyury antibody (Santa Cruz Biotechnology, Inc., catalog No. sc-20109) overnight at 4°C. After washing with PBS, the cells were incubated with Alexa Fluor 594 donkey anti-rabbit IgG (Invitrogen, catalog No. A21207) for 1 h in the dark. The cover slips were then washed with PBS and mounted with VECTASHIELD Mounting Medium with DAPI (Vector Laboratories, Burlingame, CA, USA, catalog No. H-1200). Images were visualized using an inverted microscope (ECLIPSE E600; Nikon,

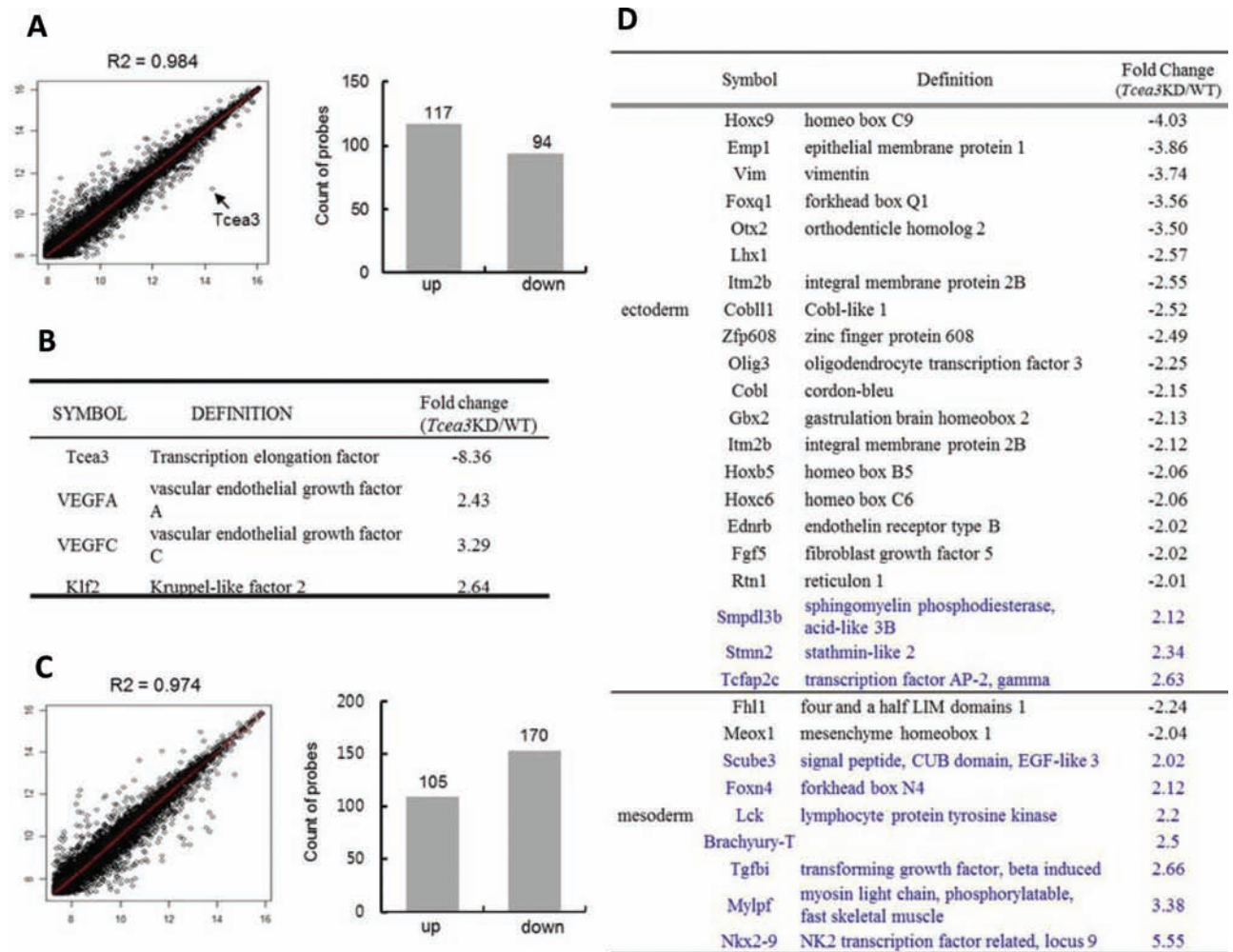
Kanagawa, Japan) and analyzed using the INFINITY2-1C software (Innerview 2.0, Lumenera, Canada).

**RESULTS**

Previously, we reported that *Tcea3* KD mESCs have an enhanced differentiation capacity (12). However, we did not address the effect of *Tcea3* on lineage-specific differentiation of mESCs. To understand the change in differentiation potential of *Tcea3* KD mESCs, we analyzed the genome-wide gene expression profiles of *Tcea3* KD mESCs under both self-renewing and early differentiating conditions.

Scatter plots of the cDNA microarray results showed that suppression of *Tcea3* did not alter the gene expression

profiles of mESCs significantly ( $R^2=0.984$ ). Among a total of 45,281 genes on the MouseWG-6 v2 Expression BeadChip (Illumina), expression of 211 genes (0.46% of the total) was significantly altered in *Tcea3* KD cells according to a Student's *t* test analysis with a 99% confidence level (upregulated: 117 genes; downregulated: 94 genes) (Fig. 1A). VEGF is a cytokine best known for inducing embryonic vasculogenesis (15). VEGFA was identified as one of 12 genes critical to vasculogenesis by systematic analysis of vascular defects in knockout mice (16), and VEGFC promotes vasculogenesis and angiogenesis in three-dimensional collagen gels (17). Interestingly, the microarray analysis of gene expression profiles of *Tcea3* KD mESCs revealed that the expression of two



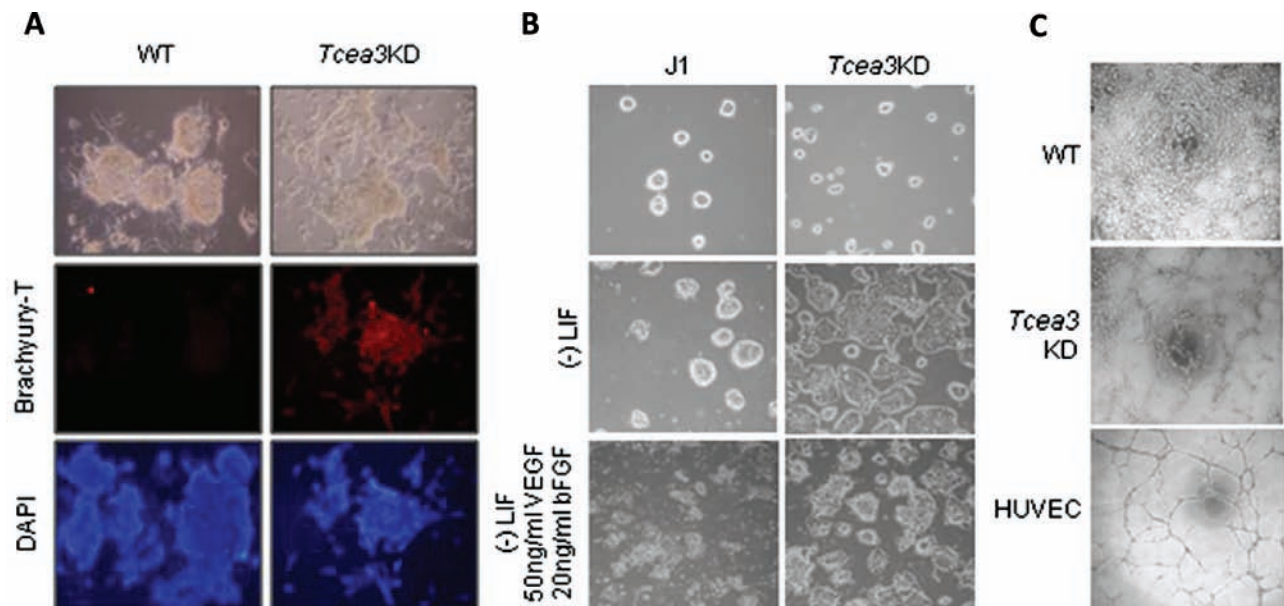
**Figure 1.** Gene expression profiles of *Tcea3* KD mESCs. (A) Scatter plots of cDNA microarray analysis of *Tcea3* KD mESCs ( $R^2$  = correlation coefficients) showing that 117 genes are activated and 94 genes are suppressed in *Tcea3* KD mESCs. (B) Panther classification of 211 genes showed that major regulators of vasculogenesis are activated in *Tcea3* KD mESCs. (C) Scatter plots of cDNA microarray analysis of differentiating *Tcea3* KD mESCs ( $R^2$  = correlation coefficients) showing that 105 genes are activated and 170 genes are suppressed in *Tcea3* KD mESCs. Differentiation of mESCs was induced by culturing in medium lacking LIF for 3 days. (D) Panther classification of 275 genes showed that 18 of 21 ectoderm lineage marker genes were suppressed, whereas 7 of 9 mesoderm lineage marker genes were activated.

isoforms of VEGF was activated in *Tcea3* KD mESCs in self-renewing culture conditions (Fig. 1B). In addition, a Klf2 transcription factor, which modulates blood vessel maturation through smooth muscle cell migration, is also activated in *Tcea3* KD mESCs (18). These results suggest that suppression of *Tcea3* expression may enhance the vascular differentiation potential of mESCs. We next induced the spontaneous differentiation of mESCs by culturing the cells in the absence of LIF for 3 days and then analyzed the gene expression profiles by microarray analysis. Like the self-renewing cells, the gene expression profiles of differentiating *Tcea3* KD cells were not significantly altered relative to mESCs ( $R^2=0.974$ ) (Fig. 1C). Among the 45,281 genes examined, 0.61% (275 genes) were significantly altered in *Tcea3* KD mESCs according to a Student's *t* test with a 99% confidence level (downregulated: 105 genes; upregulated: 170 genes). All altered genes were classified using the PANTHER classification system, which classifies genes into families and subfamilies of shared function and categorizes them by molecular function, biological process, and pathway (<http://www.pantherdb.org>). Interestingly, whereas 7 of 9 genes related to mesoderm development were activated in differentiating *Tcea3* KD mESCs, 18 of 21 ectoderm-related genes were suppressed (Fig. 1D). These results suggest that suppression of *Tcea3* expression in self-renewing ESCs leads to enhanced expression

of mesoderm lineage-related genes during differentiation of mESCs.

Based on the results of the microarray analysis of *Tcea3* KD mESCs, we next investigated whether *Tcea3* KD enhanced the differentiation capacity to mesoderm lineages. Most of the differentiating *Tcea3* KD cells that were cultured under spontaneous differentiation culture conditions (as above) for 3 days express the mesoderm marker Brachyury-T (Fig. 2A). This result is consistent with the result of the microarray analysis (Fig. 1D) and further confirms that *Tcea3* KD is prone to differentiate into mesodermal lineages.

We further confirmed the functional activity of endothelial-like cells differentiated from *Tcea3* KD cell lines. The activity was evaluated by tube formation assays after endothelial differentiation of *Tcea3* KD cell lines for 7 days. The cells were incubated in spontaneous differentiation culture medium containing VEGF (50 ng/ml) and bFGF (20 ng/ml) for 2 days (Fig. 2B). *Tcea3* KD cells showed a more differentiated cell morphology compared to wild-type (WT) mESCs in the presence of VEGF and bFGF (Fig. 2B). The cells were cultured in EGM medium for the next 5 days and then plated on Matrigel to confirm tube-forming potential (Fig. 2C). *Tcea3* KD cells formed more vessel-like structures compared to WT mESCs (Fig. 2C). HUVECs were used as a positive control for tube formation (Fig. 2C).



**Figure 2.** *Tcea3* KD mESCs have an enhanced vascular differentiation potential. (A) *Tcea3* KD and WT mESCs were induced to differentiate by culturing in medium lacking LIF for 3 days, and differentiating cells were immunostained with the mesoderm marker Brachyury-T. (B) *Tcea3* KD and WT mESCs were induced to differentiate by culturing in medium lacking LIF, in the presence or absence of VEGF and bFGF, for 3 days. Differentiating cell morphology was examined by light microscopy. (C) Tube formation of endothelial-like cells differentiated from J1 and *Tcea3* KD mESCs. After endothelial differentiation of mESCs for 7 days, the cells were plated on Matrigel to evaluate the vessel formation potential of the differentiated endothelial-like cells. More vessel-like structures were observed in endothelial-like cells differentiated from *Tcea3* KD cells compared to those from WT ESCs.

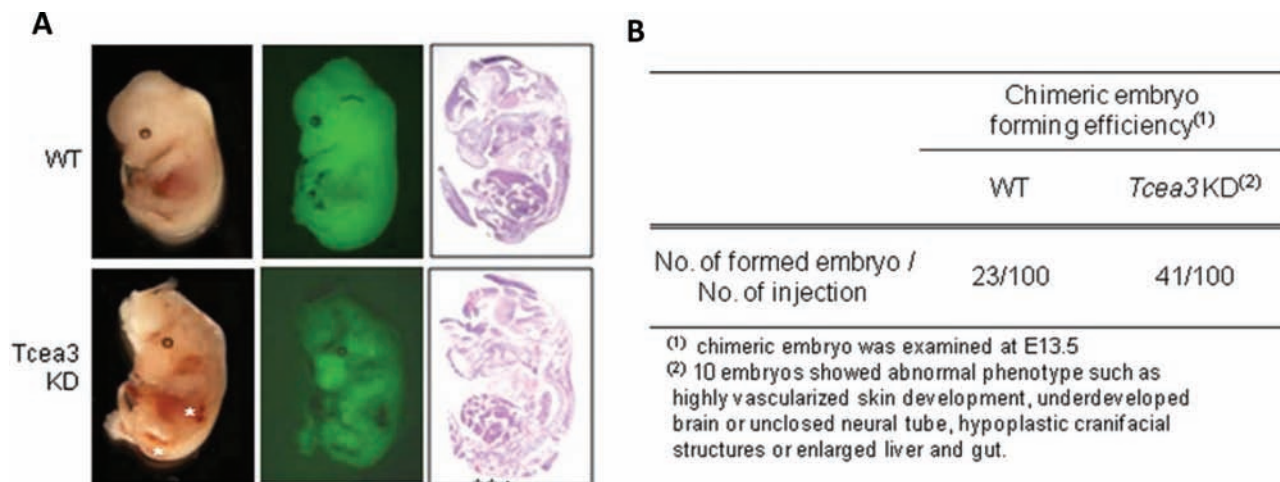
Finally, we further examined the *in vivo* developmental potential of *Tcea3* KD cells by analyzing chimeric mice. GFP-expressing mESCs were injected into mouse blastocysts, and the chimeric embryo development was examined at E13.5. Chimeric embryos were successfully generated using *Tcea3* KD as well as WT mESCs when the same numbers of GFP-expressing *Tcea3* KD or WT mESCs were injected into the cavities of blastocysts from C57BL/6 mice (Fig. 3A). These results indicate that *Tcea3* KD mESCs retain basal *in vivo* pluripotency, although the differentiation capacity of the *Tcea3* KD mESCs appeared to be significantly different during *in vitro* differentiation. Some chimeric embryos formed with *Tcea3* KD cells showed highly vascularized skin, implying that alteration of *Tcea3* expression affected *in vivo* pluripotency (Fig. 3A). In addition, other abnormal phenotypes, such as an underdeveloped brain, incomplete neural tube closure, hypoplastic cranifacial structures, and enlarged liver and gut, were observed in the chimeric embryos formed from *Tcea3* KD cells (Fig. 3B and data not shown). Taken together, these data suggest that *Tcea3* is a key regulator controlling vasculogenesis by mESCs.

### DISCUSSION

Previously, we showed that *Tcea3* regulates *Lefty1*, which is an inhibitor of Nodal-Smad2 signaling, in mESCs (12). Mesodermal differentiation markers were activated in *Tcea3* KD cells in self-renewal culture conditions, and *Tcea3* KD cells responded rapidly to external

differentiation signals, probably due to the preactivated nodal-Smad2 signaling components (5,12). Because stem cells have been intensively investigated as a potential source of cells for proangiogenic therapies, including ischemic tissues, we studied the impact of altered *Tcea3* expression on vasculogenesis by mESCs. *In vitro* and *in vivo* experiments demonstrated that suppression of *Tcea3* promotes vasculogenesis by mESCs. Genome-wide chromatin immunoprecipitation chip analysis showed that development-related genes are direct targets of phospho-Smad2 (5). Importantly, the gene expression profile suggests that the underlying mechanism of enhanced vasculogenesis by *Tcea3* KD cells is an enrichment of vasculogenesis-related genes in the ESCs under both self-renewal and differentiation conditions. Whether the expression of vasculogenesis-related genes is directly regulated by phospho-Smad2 will be important to determine in future studies. The unperturbed gene expression profiles of *Tcea3* KD cells are consistent with a previous report indicating that *Tcea3* is dispensable for self-renewal maintenance and specifically activates transcription elongation in its target genes (12).

Although more embryos developed from *Tcea3* KD chimeric blastocysts than from WT chimeric blastocysts, the *Tcea3* KD-derived embryos exhibited a number of developmental abnormalities (Fig. 3B), suggesting that modulation of gene expression by *Tcea3* is crucial for normal development. In addition to highly vascularized skin development, chimeric embryos derived from *Tcea3* KD mESCs exhibited failures in neural tube closure and



**Figure 3.** Embryos that developed from *Tcea3* KD mESCs exhibit abnormal phenotypes including highly vascularized skin. (A) Comparison of chimeric embryos generated by injecting GFP-expressing WT or *Tcea3* KD mESCs. Left panels, morphological features of representative chimeric embryos; middle panels, GFP signals from the corresponding embryos; right panels, H&E-stained embryo midsagittal sections. All embryos generated from WT cells appeared normal. By contrast, highly vascularized tissues were detected from some embryos generated from *Tcea3* KD as indicated by the asterisks. (B) The efficiency of chimeric embryo formation from blastocysts containing WT or *Tcea3* KD mESCs. Whereas all embryos generated by WT cells look normal, some chimeric embryos generated by *Tcea3* KD cells appeared developmentally abnormal.

had underdeveloped brains. These results suggest that expression level of *Tcea3* may influence brain development, as well as vasculogenesis, during embryo development. Consistent with this result, Activin and TGF- $\beta$  are reported to have effects on brain development and on the fate decisions of neural stem/progenitor cells both in vitro and in vivo (19). Therefore, elucidation of *Tcea3* activity during development will be important for understanding the process of brain formation during embryogenesis. In this study, we suggest that *Tcea3* regulates balanced lineage differentiation of mESCs, which is essential for normal development of embryo. Further studies regarding the role of *Tcea3* in vasculogenesis and angiogenesis will contribute to the understanding of molecular mechanisms of pluripotency of ESCs.

*ACKNOWLEDGMENTS:* This research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (2012M3A9C6050367, 2012RIA1A3003070). This research is also supported by the Priority Research Centers Program through the NRF funded by the Ministry of Education, Science and Technology (20120006679) and Korea Health Technology R&D project, Ministry of Health & Welfare (A091087-0911-0000200).

## REFERENCES

1. Evans MJ, Kaufman MH. Establishment in culture of pluripotential cells from mouse embryos. *Nature* 1981; 292(5819):154–156.
2. Rossant J. Stem cells and early lineage development. *Cell* 2008; 132(4):527–531.
3. Pera MF, Tam PP. Extrinsic regulation of pluripotent stem cells. *Nature* 2010; 465(7299):713–720.
4. Watabe T, Miyazono K. Roles of TGF-beta family signaling in stem cell renewal and differentiation. *Cell Res* 2009; 19(1):103–115.
5. Fei T, Zhu S, Xia K, Zhang J, Li Z, Han JD, et al. Smad2 mediates Activin/Nodal signaling in mesendoderm differentiation of mouse embryonic stem cells. *Cell Res* 2010; 20(12):1306–1318.
6. Brown S, Teo A, Pauklin S, Hannan N, Cho CH, Lim B, et al. Activin/Nodal signaling controls divergent transcriptional networks in human embryonic stem cells and in endoderm progenitors. *Stem Cells* 2011; 29(8):1176–1185.
7. Lee KL, Lim SK, Orlov YL, Yit le Y, Yang H, Ang LT, et al. Graded Nodal/Activin signaling titrates conversion of quantitative phospho-Smad2 levels into qualitative embryonic stem cell fate decisions. *PLoS Genet* 2011; 7(6):e1002130.
8. Ferguson JE, 3rd, Kelley RW, Patterson C. Mechanisms of endothelial differentiation in embryonic vasculogenesis. *Arterioscler Thromb Vasc Biol* 2005; 25(11):2246–2254.
9. Gerber HP, Malik AK, Solar GP, Sherman D, Liang XH, Meng G, et al. VEGF regulates haematopoietic stem cell survival by an internal autocrine loop mechanism. *Nature* 2002; 417(6892):954–958.
10. Ng YS, Ramsauer M, Loureiro RM, D'Amore PA. Identification of genes involved in VEGF-mediated vascular morphogenesis using embryonic stem cell-derived cystic embryoid bodies. *Lab Invest* 2004; 84(9):1209–1218.
11. Labhart P, Morgan GT. Identification of novel genes encoding transcription elongation factor TFIIS (TCEA) in vertebrates: Conservation of three distinct TFIIS isoforms in frog, mouse, and human. *Genomics* 1998; 52(3): 278–288.
12. Park KS, Cha Y, Kim CH, Ahn HJ, Kim D, Ko S, et al. Transcription elongation factor *Tcea3* regulates the pluripotent differentiation potential of mouse embryonic stem cells, via the Lefty1-Nodal-Smad2 pathway. *Stem Cells* 2013; 31(2):282–292.
13. Jirmanova L, Afanassieff M, Gobert-Gosse S, Markossian S, Savatier P. Differential contributions of ERK and PI3-kinase to the regulation of cyclin D1 expression and to the control of the G1/S transition in mouse embryonic stem cells. *Oncogene* 2002; 21(36):5515–5528.
14. Kaufman MH. Postcranial morphological features of homozygous tetraploid mouse embryos. *J Anat* 1992; 180(Pt. 3): 521–534.
15. Ferrara N. VEGF and the quest for tumour angiogenesis factors. *Nat Rev Cancer* 2002; 2(10):795–803.
16. Argraves WS, Drake CJ. Genes critical to vasculogenesis as defined by systematic analysis of vascular defects in knockout mice. *Anat Rec A Discov Mol Cell Evol Biol* 2005; 286(2):875–884.
17. Bauer SM, Bauer RJ, Liu ZJ, Chen H, Goldstein L, Velazquez OC. Vascular endothelial growth factor-C promotes vasculogenesis, angiogenesis, and collagen constriction in three-dimensional collagen gels. *J Vasc Surg* 2005; 41(4):699–707.
18. Wu J, Bohanan CS, Neumann JC, Lingrel JB. KLF2 transcription factor modulates blood vessel maturation through smooth muscle cell migration. *J Biol Chem* 2008; 283(7):3942–3950.
19. Rodriguez-Martinez G, Velasco I. Activin and TGF-beta effects on brain development and neural stem cells. *CNS Neurol Disord Drug Targets* 2012; 11(7):844–855.